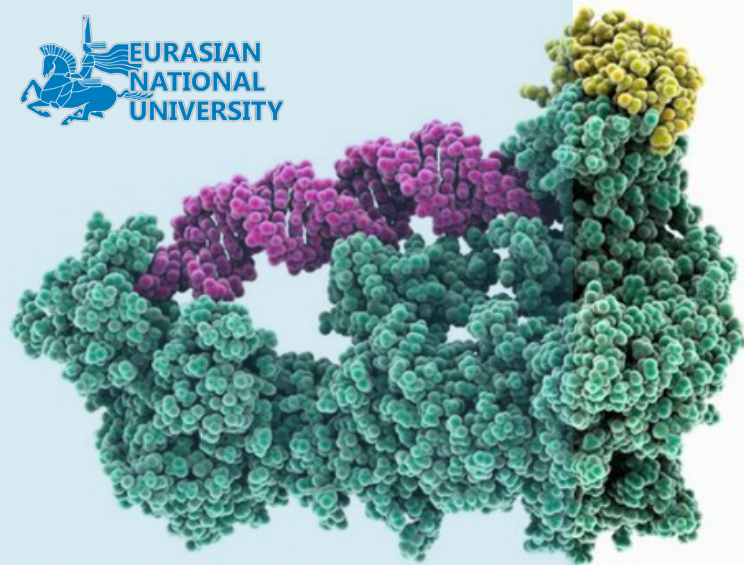


ҒЫЛЫМ ЖӘНЕ ЖОҒАРЫ БІЛІМ МИНИСТРЛІГІ
МИНИСТЕРСТВО НАУКИ И ВЫСШЕГО ОБРАЗОВАНИЯ



Л. Н. ГУМИЛЕВА АТЫНДАҒЫ
ЕУРАЗИЯ ҰЛТТЫҚ УНИВЕРСИТЕТІ

ЕВРАЗИЙСКИЙ НАЦИОНАЛЬНЫЙ
УНИВЕРСИТЕТ ИМЕНИ
Л. Н. ГУМИЛЕВА

АСТАНА, ҚАЗАҚСТАН
11 СӘУІР 2024 ЖЫЛ

АСТАНА, КАЗАХСТАН
11 АПРЕЛЯ 2024 ГОД

"ОМАРОВ ОҚУЛАРЫ: ХХІ
ҒАСЫРДЫҢ БИОЛОГИЯ ЖӘНЕ
БИОТЕХНОЛОГИЯСЫ" АТТЫ
ХАЛЫҚАРАЛЫҚ ҒЫЛЫМИ
ФОРУМНЫҢ БАЯНДАМАЛАР
ЖИНАҒЫ

СБОРНИК МАТЕРИАЛОВ
МЕЖДУНАРОДНОГО НАУЧНОГО
ФОРУМА "ОМАРОВСКИЕ ЧТЕНИЯ:
БИОЛОГИЯ И БИОТЕХНОЛОГИЯ
ХХІ ВЕКА"

УДК 57 (063)
ББК 28.0
Ж 66

Жалпы редакцияны басқарған т.ғ.д., профессор Е.Б. Сыдықов
Под редакцией д.и.н., профессора Е.Б. Сыдыкова

Редакция алқасы:
Редакционная коллегия:

Ж.К. Масалимов, А.Б. Курманбаева, Ж.А.Нурбекова, Н.Н. Иқсат.

«Омаров оқулары: ХХІ ғасыр биология және биотехнологиясы» халықаралық ғылыми форумының баяндамалар жинағы. – Астана: Л.Н. Гумилев атындағы Еуразия ұлттық университеті, 2024. – 284 б., қазақша, орысша, ағылшынша.

Сборник материалов международного научного форума «Омаровские чтения: Биология и биотехнология ХХІ века». – Астана. Евразийский национальный университет имени Л.Н. Гумилева, 2024. – 284 с., казахский, русский, английский.

ISBN 978-601-337-977-7

Жинақ «Омаров оқулары: ХХІ ғасыр биология және биотехнологиясы» атты халықаралық ғылыми форумна қатысушылардың баяндамаларымен құрастырылған. Бұл басылымда биология, биотехнология, молекулалық биология және генетиканың маңызды мәселелері қарастырылған. Жинақ ғылыми қызметкерлерге, PhD докторанттарға, магистранттарға, сәйкес мамандықтағы студенттерге арналған.

Сборник составлен по материалам, представленным участниками международного научного форума «Омаровские чтения: Биология и биотехнология ХХІ века». Издание освещает актуальные вопросы биологии, биотехнологии, молекулярной биологии и генетики. Сборник рассчитан на научных работников, PhD докторантов, магистрантов, студентов соответствующих специальностей.

ISBN 978-601-337-977-7



УДК 57
ББК 28
О-58

©Коллектив авторов, 2024
©Евразийский национальный университет имени Л.Н. Гумилева, 2024

М - маркер, 1 - 25°C, 2 - 37°C, 3 - 40°C, 4 - вирус, 5 - 37°C+вирус, 6 - 40°C+вирус
Сурет 3 - TBSV-P19 детекциясы

Берілген сурет бойынша, жоғары температуралық стресс вирустық инфекцияның төмендеуіне, яғни TBSV репликациясын тежейтіні туралы тұжырым жасауға болады. Бұл жоғары температураның өсімдіктің қорғаныс механизмдерін белсендіріп, сондай-ақ морфологиялық, физиологиялық, молекулярлық жауапты туратынын айтады.

Қаржыландыру. Берілген жұмыс AP19679597 «Біріктірілген стресс жағдайында өсімдіктердің мультижүйелік қорғанышын зерттеу» гранттық жобасы шеңберінде жасалынды.

Пайдаланылған әдебиеттер тізімі:

1. M.H. Al-Whaibi, Plant heat-shock proteins: a mini review, J. King Saud Univ. - Sci. 23 (2011) 139–150.
2. W. Wang, B. Vinocur, O. Shoseyov, A. Altman, Role of plant heat-shock proteins and molecular chaperones in the abiotic stress response, Trends Plant Sci. 9 (2004) 244–252.
3. A.L. Qu, Y.F. Ding, Q. Jiang, C. Zhu, Molecular mechanisms of the plant heat stress response, Biochem. Biophys. Res. Commun. 432 (2013) 203–207.
4. U. Lee, Complexity of the heat stress response in plants, Curr. Opin. Plant Biol. 10 (2007) 310–316.
5. A. Wahid, S. Gelani, M. Ashraf, M.R. Foolad, Heat tolerance in plants: an overview, Environ. Exp. Bot. 61 (2007) 199–223.
6. Jones RAC (2016) Future scenarios for plant virus pathogens as climate change progresses. Adv Virus Res 95:87–147. <https://doi.org/10.1016/bs.aivir.2016.02.004>
7. Jones RAC, Naidu RA (2019) Global dimensions of plant virus diseases: current status and future perspectives. Annu Rev Virol 6(1):387–409. <https://doi.org/10.1146/annurev-virology-092818-015606>
8. Lebeurier G, Hirth L (1966) Effect of elevated temperatures on the development of two strains of tobacco mosaic virus. Virology 29(3):385–395. [https://doi.org/10.1016/0042-6822\(66\)90214-5](https://doi.org/10.1016/0042-6822(66)90214-5)
9. Shamekova M. et al. Tombusvirus-based vector systems to permit over-expression of genes or that serve as sensors of antiviral RNA silencing in plants. // Virology. United States. - 2014. V. 452–453. P. 159–165.
10. Yergaliyev T.M. et al. The involvement of ROS producing aldehyde oxidase in plant response to Tombusvirus infection. // Plant Physiol Biochem. France. - 2016. V. 109. P. 36–441.

Using CRISPR cas-9 to treat cancer: A Review

Baikarayev Zhaksat Maratuly

L.N. Gumilev Eurasian National University, Astana, Kazakhstan,
zhaksatbaikaraev@gmail.com

Abstract

CRISPR-Cas9 technology has rapidly emerged as a versatile tool in genetic engineering, offering precise gene editing capabilities with unprecedented accuracy and efficiency. Originally discovered as a microbial defense mechanism(1), CRISPR-Cas9 has been ingeniously repurposed by scientists to target and modify specific genes within the human genome. This groundbreaking technology holds immense promise in significantly impacting cancer treatment by enabling the selective editing of genes associated with tumorigenesis and drug resistance,

potentially leading to more effective therapeutic interventions. In this article, we provide a comprehensive review of the current landscape of CRISPR-Cas9 applications in cancer research and treatment, highlighting its transformative potential in combating this formidable disease (2)

Introduction

Cancer is a devastating disease that claims the lives of millions of people each year. According to the World Health Organization cancer is the second leading cause of mortality in the world responsible for an estimated ten million deaths in 2020(3). At the same time, the treatment is also improving by using various genetic engineering methods such as modifying T-killer cells to enhance their ability to fight against cancer better. Traditional cancer treatments, like surgery to remove tumors, often lack specificity and come with side effects. Consequently, contemporary oncology is prioritizing the development of therapies that are more targeted and safer. (4,5) This shift has led to the emergence of other techniques. Furthermore, the advent of precise gene editing tools represents a milestone, opening doors to treatment modalities that can directly target the genes driving the uncontrolled growth and survival of cancer cells. But curing cancer is challenging due to the complex mechanisms and because of the diversity of cancer types and their ability to adapt to any environment it is hard to detect mutated genes and finding a universal cure is still impossible.(6) But CRISPR (Clustered Regularly Interspaced Short Palindromic Repeats), first discovered in *E. coli* in 1987, fundamentally changed our understanding of DNA and cancer therapy. Unlike other genetic engineering techniques, the use of CRISPR-Cas9 has improved throughout the years. With its unparalleled precision, CRISPR-Cas9 holds the potential to revolutionize cancer therapy by enabling targeted gene editing to eradicate cancerous cells while sparing healthy tissue(7). Because of the specificity and efficacy CRISPR-Cas9 become a significant advancement in biochemistry. In this article, we will review CRISPR-Cas9 and its applications in cancer therapy and research also basic mechanisms of this technique

Mechanisms of CRISPR

For centuries we did not know about how short repeat sequences work. But in 2005 scientists found out that these sequences are part of an immune system in bacteria(8). Thus, they concluded that this CRISPR/Cas9 technology originated from a fascinating immune defense mechanism observed in bacteria and archaea, providing them with protection against invading nucleic acids like viruses and phages. This system, known as Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) along with the Cas9 enzyme, has gained significant attention in genetic engineering. Typically, CRISPR/Cas systems are classified into three main types, each comprising various subgroups. Among these, the type II CRISPR/Cas system is most employed for gene editing. It consists of three key components: Cas9, CRISPR RNA (crRNA), and transactivating crRNA (tracrRNA). The crRNA and tracrRNA molecules join to form a duplex structure called guide RNA (gRNA)(9). To streamline the process of genome engineering, this gRNA can be replaced by a synthetic fused chimeric single gRNA (sgRNA), making CRISPR/Cas9 technology more user-friendly and accessible. In the realm of genetic engineering, the single guide RNA (sgRNA) plays a pivotal role(10). Crafted with precision, it boasts a distinctive twenty base-pair (bp) sequence meticulously tailored to complement the target DNA site. For compatibility with the Cas9 protein, the sequence must be followed by a concise DNA segment called “protospacer-adjacent motif” (PAM) which is a short DNA sequence, usually 2 to 6 nucleotides, located near the target DNA sequence that Cas9 needs to cut. Upon expression within the cell, the sgRNA joins forces with the Cas9 nuclease, forming a formidable ribonucleoprotein (RNP) complex(11). Guided by the sgRNA, this dynamic duo navigates to the designated target DNA site with remarkable accuracy. The incision, occurring within the protospacer, occurs with surgical precision, precisely three nucleotides upstream of the PAM, yielding blunt ends. Facilitated by the RuvC and HNH active-site motifs of Cas9, this cleavage simultaneously targets both the (–) and (+) DNA strands. Subsequently, the cell's repair machinery springs into action, using one of two primary mechanisms—homology-directed repair (HDR) or non-homologous end joining (NHEJ). Homology-directed repair (HDR): This process

uses a donor DNA template to accurately repair the DNA double-strand break (DSB)(12). It is used for precise genome editing, such as introducing specific sequences or mutations. Non-homologous end joining (NHEJ): This mechanism is more common but less precise. It tends to insert or delete nucleotides at the DSB site, often causing frameshift mutations. It is useful for inducing gene knockouts. Also, this kind of mutation can be dangerous because of the randomness of the repair mechanism(12,13).

Overall, CRISPR/Cas9 holds a big promise in the field of genetic engineering. It is much easier to use, and design compared to older methods (ZFNs and TALENs)(14). Traditional methods rely on engineering proteins for each target gene. But CRISPR/Cas9 uses sgRNA instead. The RNA acts like a search term, providing Cas protein with the right spot in the DNA. Cas9 then makes a clean-cut at that location. Allowing scientists to introduce precise changes.

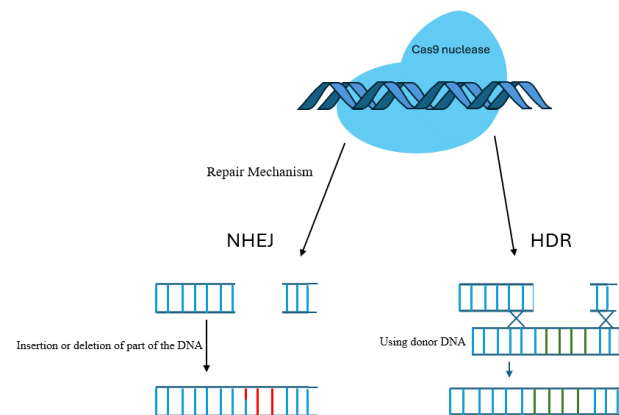


Fig 1. Two possible pathways for repairing a double-strand break in DNA: non-homologous end joining (NHEJ) and homology-directed repair (HDR).

CRISPR/Cas9 in cancer therapy

Finding a cure for cancer is still a complex problem. Despite improvements, there are still obstacles to overcome before CRISPR-Cas9 can be widely adopted in cancer therapy. One major challenge is delivering the CRISPR-Cas9 system accurately and efficiently into the target cells. In the context of cancer therapy, CRISPR-Cas9 offers diverse benefits. One important use is its potential to disable or change oncogenes, which are genes that promote cancer growth(15). By targeting and modifying these genes, CRISPR-Cas9 has the potential to slow down or even halt tumor growth. Earlier gene editing tools like TALENs and ZFNs were used for cancer treatment, but they may not have been as specific in targeting the exact epigenetic modifications associated with the disease(16). However, there has been a significant advancement in the field of immunology. Scientists introduced genetically engineered T-killer cells which are called CAR-T cells. Using the strength of the patient's own immune system, CAR T-cell therapy is a groundbreaking method of treating cancer. The creation of chimeric antigen receptors (CARs) is crucial to this therapy(17). Chimeric antigen receptors (CARs) are the workhorses of CAR T-cell therapy. They are engineered proteins that combine functionalities from various parts of the immune system to give T cells the ability to recognize and target specific cancer cells(18). CAR-T cell therapy has shown great promise in treating blood cancers due to the ability of cancer cells to move freely throughout the bloodstream(5). However, solid tumors pose a challenge to therapy as the dense tumor microenvironment can prevent CAR-T cells from reaching all cancer cells(19) (20) Developing effective CAR-T cells targeting all cancer cells is a complex task due to the wide range of surface proteins in solid tumors(21). CAR T-cell therapies can sometimes lead to a condition called T-cell exhaustion, where the modified T cells gradually lose their

effectiveness over time(22). Additionally, these engineered T cells may not remain in the body for an extended period, which can limit their long-term impact on cancer treatment. But, with the help of CRISPR/Cas9, we can modify these CAR-T cells to be more persistent and potent(23). We can use CRISPR-Cas9 to eliminate genes that make CAR T cells less effective, such as genes that cause T-cell exhaustion, or insert genes that make CAR T cells more effective, such as genes that help them to survive and multiply. CRISPR can target genes like PD-1 or CTLA-4, which function as brakes on the immune system, allowing CAR T cells to function for longer periods (24).

Scientists can use CRISPR to modify an animal's genome, including inserting, deleting, or changing specific genes. GEMMs (Genetically Engineered Mouse Models) are a valuable tool in cancer research because they allow scientists to study the development and progression of cancer in a living organism. By creating mice with mutations that are known to cause cancer in humans, researchers can gain insights into the biology of cancer and test potential new therapies(25).

Conclusion

CRISPR-Cas9 technology has emerged as a revolutionary tool with immense potential for cancer treatment. Its exceptional precision allows for targeted editing of genes associated with tumorigenesis and drug resistance, paving the way for more effective therapeutic interventions. While challenges remain in delivery methods and ensuring long-term efficacy, the ongoing advancements in CRISPR-Cas9, particularly its use in engineering CAR-T cells, offer a ray of hope for a future with more successful and personalized cancer therapies. However, it is crucial to acknowledge the ethical considerations surrounding this powerful technology as we move forward in its development and application.

References

1. Zhang H, Qin C, An C, Zheng X, Wen S, Chen W, et al. Application of the CRISPR/Cas9-based gene editing technique in basic research, diagnosis, and therapy of cancer. *Mol Cancer*. 2021 Dec 1;20(1):126.
2. Hsu PD, Lander ES, Zhang F. Development and applications of CRISPR-Cas9 for genome engineering. *Cell*. 2014 Jun 5;157(6):1262–78.
3. Ferlay J, Colombet M, Soerjomataram I, Parkin DM, Piñeros M, Znaor A, et al. Cancer statistics for the year 2020: An overview. *Int J Cancer*. 2021 Aug 15;149(4):778–89.
4. Dimitri A, Herbst F, Fraietta JA. Engineering the next-generation of CAR T-cells with CRISPR-Cas9 gene editing. *Mol Cancer*. 2022 Mar 18;21(1):78.
5. June CH, O'Connor RS, Kawalekar OU, Ghassemi S, Milone MC. CAR T cell immunotherapy for human cancer. *Science* (1979). 2018 Mar 23;359(6382):1361–5.
6. Akram F, Ikram ul Haq, Ahmed Z, Khan H, Ali MS. CRISPR-Cas9, A Promising Therapeutic Tool for Cancer Therapy: A Review. *Protein Pept Lett*. 2020 Nov 2;27(10):931–44.
7. Ran FA, Hsu PD, Wright J, Agarwala V, Scott DA, Zhang F. Genome engineering using the CRISPR-Cas9 system. *Nat Protoc*. 2013 Nov 24;8(11):2281–308.
8. Barrangou R, Fremaux C, Deveau H, Richards M, Boyaval P, Moineau S, et al. CRISPR provides acquired resistance against viruses in prokaryotes. *Science*. 2007 Mar 23;315(5819):1709–12.
9. Manghwar H, Li B, Ding X, Hussain A, Lindsey K, Zhang X, et al. CRISPR/Cas Systems in Genome Editing: Methodologies and Tools for sgRNA Design, Off-Target Evaluation, and Strategies to Mitigate Off-Target Effects. *Advanced Science*. 2020 Mar 6;7(6).
10. Zhang F, Wen Y, Guo X. CRISPR/Cas9 for genome editing: progress, implications and challenges. *Hum Mol Genet*. 2014 Sep 15;23(R1):R40–6.
11. Sternberg SH, Doudna JA. Expanding the Biologist's Toolkit with CRISPR-Cas9. Vol. 58, *Molecular Cell*. Cell Press; 2015. p. 568–74.
12. Torgovnick A, Schumacher B. DNA repair mechanisms in cancer development and therapy. *Front Genet*. 2015 Apr 23;6.

13. Sánchez-Rivera FJ, Jacks T. Applications of the CRISPR–Cas9 system in cancer biology. *Nat Rev Cancer*. 2015 Jul 4;15(7):387–93.
14. Jiang F, Doudna JA. CRISPR–Cas9 Structures and Mechanisms. *Annu Rev Biophys*. 2017 May 22;46(1):505–29.
15. Jiang C, Meng L, Yang B, Luo X. Application of *CRISPR/Cas9* gene editing technique in the study of cancer treatment. *Clin Genet*. 2020 Jan 10;97(1):73–88.
16. Shankar S, Sreekumar A, Prasad D, Das A V., Pillai MR. Genome editing of oncogenes with ZFNs and TALENs: caveats in nuclease design. *Cancer Cell Int*. 2018 Dec 22;18(1):169.
17. June CH, O’Connor RS, Kawalekar OU, Ghassemi S, Milone MC. CAR T cell immunotherapy for human cancer. *Science* (1979). 2018 Mar 23;359(6382):1361–5.
18. Huang R, Li X, He Y, Zhu W, Gao L, Liu Y, et al. Recent advances in CAR-T cell engineering. *J Hematol Oncol*. 2020 Dec 2;13(1):86.
19. Newick K, O’Brien S, Moon E, Albelda SM. CAR T Cell Therapy for Solid Tumors. *Annu Rev Med*. 2017 Jan 14;68(1):139–52.
20. Martinez-Lage M, Puig-Serra P, Menendez P, Torres-Ruiz R, Rodriguez-Perales S. CRISPR/Cas9 for Cancer Therapy: Hopes and Challenges. *Biomedicines*. 2018 Nov 12;6(4):105.
21. Shah NN, Fry TJ. Mechanisms of resistance to CAR T cell therapy. *Nat Rev Clin Oncol*. 2019 Mar 5;
22. Wang Z, Wu Z, Liu Y, Han W. New development in CAR-T cell therapy. *J Hematol Oncol*. 2017 Dec 21;10(1):53.
23. Sadeqi Nezhad M, Yazdanifar M, Abdollahpour-Alitappeh M, Sattari A, Seifalian A, Bagheri N. Strengthening the CAR-T cell therapeutic application using CRISPR/Cas9 technology. *Biotechnol Bioeng*. 2021 Oct 21;118(10):3691–705.
24. Agarwal S, Wellhausen N, Levine BL, June CH. Production of Human CRISPR-Engineered CAR-T Cells. *Journal of Visualized Experiments*. 2021 Mar 15;(169).
25. Kersten K, de Visser KE, van Miltenburg MH, Jonkers J. Genetically engineered mouse models in oncology research and cancer medicine. *EMBO Mol Med*. 2017 Feb 27;9(2):137–53.

УДК 574.24

**БАТЫС ҚАЗАҚСТАН ОБЛЫСЫНДА КЕЗДЕСЕТІН ИКСОД
КЕНЕЛЕРІНІҢ (*IXODIDAE, PARASITIFORMES*) ҚОРЕКТЕНУ ЕРЕКШЕЛІКТЕРІ**

Төлеуова Раушангуль Нурлановна

Абай атындағы Қазақ ұлттық педагогикалық университеті, Алматы, Қазақстан

Raushan_gul_85@mail.ru

Ғылыми жетекшісі- Есімов Б.К.

Иксодид кенелері ұзақ уақыт қоректенетін уақытша эктопаразиттердің экологиялық тобына жатады [1], сондықтан өмірлік цикл иксодид төрт кезеңнен тұрады: жұмыртқа, личинка, нимфа және имаго. Личинкалар мен нимфаларда қанмен қаныққаннан кейін балқу пайда болады, ал ересек аналықтар қанның көп мөлшерін ішеді, олардың массасы бірнеше есе артады.

Жердегі артроподтардың паразиттену түрлерін зерттеу барысында В. Н. Беклемишев (1970) түрдің тіршілік схемасы туралы тұжырымдама жасады [2].

Соңғысы түрдің өмірлік циклін, иесінің денесімен де, қоршаған ортамен де қарым-қатынастағы барлық кезеңдерінің ерекшеліктерін қамтиды.

Кейіннен Ю. С. Балашов бұл жіктеуді паразиттік-ие қатынастар негізінде кеңейтті [3].